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UTILITY PATENT APPLICATION TRANSMITTAL
(new nonprovisional applications under 37 CFR 1.53(b))

Transmitted herewith for filing is the patent application of:

INVENTOR(S): Thomas M. BEHR and David M. GOLDENBERG

TITLE: METHODS FOR REDUCED RENAL UPTAKE OF PROTEIN CONJUGATES

In connection with this application, the following are enclosed:

APPLICATION ELEMENTS:

☒ Specification - 42 TOTAL PAGES

(preferred arrangement:)

- Descriptive Title of the Invention
- Cross Reference to Related Applications
- Statement Regard Fed sponsored R&D
- Reference to Microfiche Appendix
- Background of the Invention
- Brief Summary of the Invention
- Brief Description of the Drawings (if filed)
- Detailed Description
- Claim(s)
- Abstract of the Disclosure

☒ Drawings - Total Sheets 7

☐ Declaration and Power of Attorney - Total Sheets

☐ Newly executed (original or copy)

☐ Copy from a prior application (37 CFR 1.63(d))

(relates to continuation/divisional boxes completed) - NOTE: Box below

☐ DELETION OF INVENTOR(S) - Signed statement attached deleting inventor(s)
named in the prior application, see 37 CFR 1.63(d)(2) and 1.33(b).

☐ Incorporation By Reference (useable if copy of prior application
Declaration being submitted)

The entire disclosure of the prior application, from which a COPY of the
oath or declaration is supplied as noted above, is considered as being
part of the disclosure of the accompanying application and is hereby
incorporated by reference therein.

☐ Microfiche Computer Program (Appendix)

☐ Nucleotide and/or Amino Acid Sequence Submission (if applicable, all necessary)

☐ Computer Readable Copy

☐ Paper Copy (identical to computer copy)

☐ Statement verifying identify of above copies

ACCOMPANYING APPLICATION PARTS

☐ Assignment Papers (cover sheet & document(s))

☐ 37 CFR 3.73(b) Statement (when there is an assignee)

☐ English Translation Document (if applicable)

☐ Information Disclosure Statement (IDS) with PTO-1449. ☐ Copies of IDS Citations

☐ Preliminary Amendment

☒ Return Receipt Postcard (MPEP 503)

☒ Small Entity Statement(s)

☒ Statement filed in prior application, status still proper and desired.

☐ Certified Copy of Priority Document(s) with Claim of Priority
(if foreign priority is claimed).

☐ OTHER:

If a CONTINUING APPLICATION, check appropriate box and supply the requisite information:

☐ Continuation ☐ Divisional ☒ Continuation-in-part (CIP)
of prior application Serial No. 08/407,899, filed March 21, 1995 (U.S. Patent No. 5,843,849, issued December 1, 1998).

☒ Amend the specification by inserting before the first line the following sentence:
--This application is a continuation-in-part of U.S. Application Serial No. 08/407,899, filed March 21, 1995 (U.S. Patent No. 5,843,849, issued December 1, 1998).--

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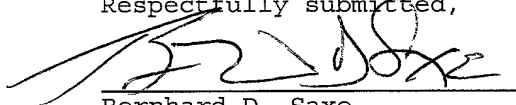
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Independent Claims	2 - 3 =	0	x \$78 (x \$39)	0
Multiple Dependent Claims			\$260 (\$135)	0
Assignment Recording Fee per property			\$40	0
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Respectfully submitted,



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Date: November 30, 1998

Docket No.: 018734/0161

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METHODS FOR REDUCED RENAL UPTAKE OF PROTEIN CONJUGATES

Background of the Invention

The invention provides methods of reducing renal uptake and retention of protein conjugates during therapy and imaging.

In particular, one aspect of the invention relates to a method for reducing renal uptake of monoclonal antibody fragments used for radioimmuno-diagnosis (RAID), immunotherapy, and radioimmunotherapy (RAIT). Recent approaches in diagnosis and therapy of disease have focused on methods of delivering radionuclides and other diagnostics or therapeutic agents to a preselected tissue by attaching the radionuclide or agent to a targeting protein which will then localize to that tissue. For example, numerous clinical studies have demonstrated the utility of radiolabeled antibodies for the radioimmunodetection of disease. Antibody fragments, such as Fab', Fab, F(ab')₂ and F(ab)₂, have faster targeting kinetics than intact immunoglobulin, and are therefore particularly useful for RAID applications. Other advantages of antibody fragments include a much lower occurrence of human immune responses than is found with intact IgG molecules.

A major drawback to the use of protein conjugates and radiolabeled antibody fragments for imaging and therapy is the relatively high uptake and retention of cytotoxic agents and radioactivity in the kidney. For example, this phenomenon can impede diagnostic accuracy, especially in the periaortic and epigastric region, and particularly when intracellularly retained isotopes, such as ¹¹¹In or ^{99m}Tc are used. Thus, when ^{99m}Tc-labeled Fab' fragments are used, renal uptakes of up to 25 % of the injected dose within 24 h have been reported.

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The maximum tolerated dose for a kidney in external beam radiation is estimated to be about 2000 cGy. Above that threshold the risk of radiation nephritis with subsequent scarring of glomeruli, nephrotic syndrome and renal insufficiency rises considerably. Furthermore, nephrotoxicity and renal toxicity are also dose-limiting due to renal retention of proteins coupled to cytotoxic agents. It is likely, therefore that in therapeutic methods using proteins and antibody fragments labeled with intracellularly retained radiometals (e.g., ^{90}Y , $^{168/188}\text{Re}$, ^{67}Cu , ^{177}Lu) and in treatments involving proteins coupled to cytotoxic agents, the kidney may become the dose-limiting organ.

Renal uptake of peptides and small proteins is thought to occur via glomerular filtration of molecules smaller than 60 kD, with subsequent tubular reabsorption for lysosomal degradation. When radioisotopes such as iodine are liberated by this degradative process they are released quickly from the cell, but radiometals are retained by binding to ubiquitous intracellular proteins with high affinity for metal ions.

Basic amino acids, such as L-lysine and L-arginine, have been shown to induce functional proteinuria when administered in high dose. Morgenson *et al.*, *Scan. J. Clin. Lab. Invest.* 37:477 (1977). Two previous studies have suggested L-lysine can be effective in decreasing kidney uptake of radiolabeled peptides. Continuous infusion of an amino acid solution containing L-lysine and L-arginine was shown to reduce renal uptake of the ^{111}In -labeled somatostatin analogue octreotide in humans. Hammond *et al.* *Brit. J. Cancer* 67:1437 (1993). Repeated intraperitoneal injections of high doses of L-lysine also reduced renal uptake of ^{111}In -labeled Fab' fragments in BALB/c mice. Pimm *et al.* *Eur. J. Nucl. Med.* 21:663 (1994).

The requirement for either continuous infusion or repeated injection of amino acids in the methods described by Hammond *et al.* and by Pimm *et al.* would cause substantial inconvenience and increased costs in a clinical setting. In addition, the most

clinically efficacious dosages of amino acids approach the maximum levels that can be physiologically tolerated before toxicity is observed.

It is apparent, therefore that more convenient methods for reducing renal retention of radiolabeled proteins or antibody fragments and protein conjugates of cytotoxic agents are greatly to be desired. Methods that are less toxic, and that require fewer and lower dosages are particularly desirable.

Summary of the Invention

It is therefore an object of the present invention to provide methods that greatly reduce kidney uptake of protein conjugates, including antibody fragment conjugates.

In accomplishing the foregoing object of the invention, there has been provided, in accordance with one aspect of the invention, a method of reducing kidney uptake of protein conjugates including antibody fragment conjugates in a patient during diagnosis or therapy, comprising administering to the patient an effective amount of one or more members compounds selected from the group consisting of D-lysine, poly-D-lysine, and poly-L-lysine, or a pharmaceutically acceptable salt or carboxyl derivative thereof. In a preferred embodiment the poly-D-lysine and the poly-L-lysine each have a molecular weight of 15-30 kDa. In accordance with another aspect of the invention is a radiolabeled conjugate. In another preferred embodiment the radiolabel is an imaging isotope and the procedure is radioimmunodiagnosis. In another preferred embodiment the conjugate is a cytotoxic agent and the procedure is immunotherapy.

In another preferred embodiment the radiolabel is an imaging isotope, and the procedure is radioimmunodiagnosis. In another preferred embodiment the conjugate comprises a cytotoxic agent and the procedure is immunotherapy.

In yet another preferred embodiment, the protein conjugate radiolabel is a therapeutic isotope.

In accordance with another aspect of the invention there have been provided methods wherein D-lysine, poly-D-lysine, or poly-L-lysine is administered to said patient.

In accordance with yet another aspect of the invention, there has been provided a method wherein a mixture of at least two of compounds selected from D-lysine, poly-D-lysine, and poly-L-lysine is administered to said patient.

There has been provided, in accordance with still another aspect of the invention, a method wherein a compound selected from the group consisting of D-lysine, poly-D-lysine, and poly-L-lysine is parenterally administered to a patient in a physiologically acceptable aqueous solution. In a preferred embodiment the solution is administered by continuous infusion. In another preferred embodiment the solution is administered by means of at least one injection of a bolus of said solution

In accordance with yet another aspect of the invention, there has been provided a method wherein a compound selected from the group consisting of D-lysine, poly-D-lysine, and poly-L-lysine is orally administered to a patient. In yet another preferred embodiment a physiologically acceptable aqueous solution is administered by means of at least one injection of a bolus of said solution followed by oral administration in a physiologically acceptable carrier.

In accordance with yet another aspect of the invention, there has been provided a method for the reduction in renal uptake and retention of radiolabeled protein conjugates and cytotoxic agent conjugates during treatment with targeting protein conjugates.

Other objects, features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become

apparent to those skilled in the art from this detailed description.

Brief Description of the Drawings

Figure 1 shows the kidney uptake kinetics and organ distribution of ^{99m}Tc -Fab' NP-4 in BALB/c mice.

Figure 2 shows the dose-effect relationship between L-lysine hydrochloride, administered intraperitoneally in hourly intervals, and the kidney uptake of ^{99m}Tc -labeled Fab' fragments of the anti-CEA MAb NP-4 in BALB/c mice, after 4h (upper graph) or 24h (lower graph) post-injection.

Figure 3 shows the effect of L-lysine treatment on kidney uptake of directly ^{188}Re -labeled anti-CSAp Mu-9 Fab' in nude mice bearing GW39 colonic cancer xenografts.

Figure 4 shows the effect of L-lysine treatment on kidney uptake of ^{111}In (upper graph) and ^{88}Y -Bz-DTPA (lower graph) labeled Fab' MN-14 in GW39 bearing nude mice colonic cancer xenografts.

Figure 5 shows the effect of the frequency of the intraperitoneal administration of lysine, poly-L-lysine and mixtures thereof on the kidney uptake of ^{99m}Tc -Fab' NP-4.

Figure 6 shows a time course of the effect of L-lysine on reduction of kidney uptake of ^{88}Y and ^{111}In -labeled F(ab)₂ fragments of the anti-CEA antibody MN-14.

Figure 7 shows the effects of a commercially available amino acid solution (containing 1.75 g of L-lysine) on kidney uptake in five patients undergoing RAID studies with ^{99m}Tc -Fab' fragments of the anti-CEA MAbs F023C5 and NP-4. Control patients were given an equal volume of saline.

Detailed Description

The present invention provides a means of achieving a significant reduction in the kidney retention of protein conjugates during diagnosis or therapy, for example, the methods reduce kidney retention of antibody fragment conjugates during RAID or immunotherapy. The reduction is achieved by administration to patients of solutions containing D-lysine alone, or lysine plus poly-lysine, prior to the procedure. Additional doses of the solutions may also be administered after delivery of the protein conjugate.

As used herein, a "significant" reduction in renal uptake and retention of radioisotope in a patient treated with protein conjugates or antibody fragment conjugates during a diagnostic procedure means a reduction at the time of imaging by at least approximately a factor of 2, more preferably a factor of approximately 3 or greater, relative to a procedure in which the solutions of the invention are not used. Another measure of "significant" reduction in renal uptake and retention of radioisotope is the ability to clearly detect and image a tumor or infectious lesion that is otherwise obscured by high background radiation in the vicinity of the kidney when the solutions of the invention are not used, especially at short imaging times of, e.g. 1-5 hours. In general, the reduction will be most pronounced at short imaging times, affording real advantages to the clinician.

Similarly, a "significant" reduction in renal uptake and retention of radioisotope or cytotoxic agent during therapy means that a dose of protein conjugate, such as an antibody fragment conjugate can be used that is at least approximately 2-3 fold times higher than could otherwise be used without risk of kidney damage.

The methods of the present invention especially are useful in reducing kidney retention of radiolabeled protein conjugates and cytotoxic agent protein conjugates in patients

undergoing treatment with targeting protein conjugates that preferentially bind to cells and tissues which are associated with a disease state. By killing these cells or tissues, the protein conjugates alleviate the disease state.

Alternatively, radiolabeled targeting proteins provide a diagnostic tool when the radioisotope in the radiolabeled protein conjugate is an imaging isotope and the targeting protein preferentially binds to cells and tissues which are associated with a disease state. Binding occurs to complementary molecules and structures associated with or expressed on the surface of the diseased cells or tissue, which preferably are not associated with or expressed on the surface of healthy cells. More typically the complementary moieties will be present on healthy cells, but to a lesser extent than is observed in the disease state.

The present invention also relates to target protein conjugates which react with a substance associated with a targeted cell which internalizes the conjugate into the cell. Targeting protein conjugates which are internalized into a cell are extremely potent cytotoxic agents for cells that are responsible for many human diseases. Because of their high activity, radiolabels and cytotoxic agents have been attached to proteins which react with a substance associated with a targeted cell in order to form cytotoxic agents which specifically bind to target cells. These protein conjugates are, therefore, very useful in therapy for cancer and infections caused by viruses, parasites, and other infectious organisms.

The conjugate comprises a cytotoxin or radiolabel and a protein which reacts with a substance associated with a targeted cell or pathogen. The substance internalizes the conjugate into the cell or into the cytoplasm of the pathogen and the conjugate subsequently kills the cell or pathogen.

A major drawback to the use of targeting protein conjugates for imaging and therapy is the relatively high uptake and retention of cytotoxic agents and radioactivity in the kidney. The present invention provides methods for reducing renal uptake and retention of protein conjugates such that a

dose of targeting protein conjugate can be administered that is at least approximately 2-3 fold times higher than could otherwise be used without risk of kidney damage.

In a preferred embodiment, the present invention is particularly useful for reducing kidney reuptake and retention of targeting protein conjugates during treatment with protein conjugates of ribonucleases such as an onconase or protein conjugates of recombinant forms thereof. Onconase is a non-mammalian ribonuclease (RNase) with a molecular weight of 12,000 that can be purified from *Rana pipiens* oocytes and early embryos. Onconase causes potent inhibition of protein synthesis in a rabbit reticulocyte lysate (IC_{50} 10^{-11} M) and when microinjected into *Xenopus* oocytes (IC_{50} 10^{-10} M). Unlike other members of the RNase A superfamily, onconase does not degrade oocyte rRNA. Upon binding to the cell surface receptors of sensitive cells and its cytosolic internalization, onconase causes cell death as a result of potent protein synthesis inhibition by a mechanism involving inactivation of cellular RNA. Onconase is not inhibited by mammalian placental ribonuclease inhibitor and this may explain Onconase's enhanced cytotoxicity when compared to the mammalian enzymes.

Animal toxicology studies show that Onconase displays a predictable, dose-dependent and reversible toxicity in both rats (dose range 0.01-0.02 mg/kg) and dogs (0.005-0.15 mg/kg). Mice inoculated with the aggressive M109 Madison lung carcinoma and treated with both daily and weekly schedules of intraperitoneally-administered Onconase, showed significantly prolonged survival. Most striking results were seen in a group of mice treated with a weekly schedule of Onconase in which six of eighteen animals survived long-term and were apparently cured of cancer.

Onconase has been shown in clinical trials to have anti-tumor activity against a variety of solid tumors. In this regard it has been used both alone and combined with other anti-tumor agents such as tamoxifen, e.g., when treating patients with pancreatic cancer. In a Phase I study, patients suffering from a variety of relapsing and resistant tumors

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were treated intravenously with Onconase. The maximum tolerated dose, mainly due to renal toxicity, was found to be 960 $\mu\text{g}/\text{m}^2$. When used as an anti-tumor agent, Onconase can be conjugated to a marker which targets it to a specific cell type.

Onconase thus has many desirable characteristics, including small size, animal origin, and anti-tumor effects *in vitro* and *in vivo*. It is well-tolerated and refractory to human RNase inhibitors. Accordingly, it is therapeutically useful to provide treatment with targeting protein-Onconase conjugates.

However, renal toxicity is potentially indicated during such treatment due to the cytotoxicity of such conjugates and the small size of the Onconase molecules, which can result in conjugates that are readily filterable by the kidney. Thus, the methods of the present invention are particularly useful in reducing renal toxicity in such treatments.

Another promising approach to therapy and diagnosis involves a pretargeting method comprising directing antibody fragment or protein conjugates to disease tissues, wherein the antibody or antibody fragment can target a diagnostic agent or therapeutic agent to a predetermined site. One method involves use of bi-specific monoclonal antibodies (bsMAbs) having at least one arm that is reactive against a targeted diseased tissue and at least one other arm that is reactive against a low molecular weight protein hapten conjugate that is either radiolabeled or conjugated with a cytotoxic agent. In this methodology, a bsMAb is administered and allowed to localize to target, and to clear normal tissue. Some time later, a radiolabeled low molecular weight hapten or a hapten conjugated with a cytotoxic agent is given, which is recognized by the second specificity of the bsMAb, and therefore localizes to the original target.

It will be readily appreciated by those skilled in the art that the methods of the present invention are particularly useful in this methodology since renal toxicity and nephrotoxicity of the conjugated hapten can be significantly

reduced or eliminated since renal uptake and retention of conjugated haptens is reduced.

Some radioisotopes that are preferred for RAID applications, such as technetium-99m, have short half-lives (for example, $t_{1/2}$ of Tc-99m is six hours). This means that rapid targeting of an antibody with one of these isotopes is desirable. Antibody fragments such as $F(ab')_2$ and $F(ab)_2$, and especially Fab, Fab', show more rapid targeting kinetics than whole immunoglobulin, and are also associated with a much lower incidence of human anti-murine antibody (HAMA) immune responses. Therefore, they are preferred for RAID applications using isotopes such as Tc-99m.

For immunotherapy applications, whole antibody or bivalent antibody fragments such as $F(ab')_2$ and $F(ab)_2$ are often preferred due to their higher binding constants. Whole antibodies usually are effectively cleared through the liver, and do not pose a significant problem of kidney toxicity. $F(ab')_2$ and $F(ab)_2$ antibody fragments, however, though too large to be effectively filtered through the glomerular basal membrane, nevertheless can still cause significant problems with kidney retention and toxicity during immunotherapy or RAIT.

Previous workers have shown that a continuously infused solution of basic L-amino acids reduced kidney retention of radioactivity in patients being treated with a radiolabeled peptide. See Hammond et al., *supra*. It also has been shown that repeated high doses of L-lysine can reduce the level of retention of radioactivity in the kidneys of mice treated with radiolabeled antibody fragments. See Mimm et al., *supra*.

In the methods described in the prior work, however, the maximum achievable reduction in kidney retention is limited by the dosage of basic amino acids that can be tolerated without causing toxicity. This toxicity is presumably due to effects of the naturally-occurring L-amino acids on amino acid metabolism. Another drawback to the previous work was that the basic amino

acids had to be given repeatedly or continuously to achieve the desired effects.

The present inventors have shown that D-lysine is highly effective in reducing kidney retention of radioactivity in subjects that are treated with radiolabeled proteins such as antibody fragments. D-lysine is highly effective in reducing kidney retention of cytotoxic agents in subjects that are treated with protein and antibody fragment conjugates of cytotoxic agents. D-Lysine does not occur naturally in humans or animals and is believed to be metabolically inert, thereby reducing the risk of toxic side-effects associated with the use of L-lysine. Mixtures of D- and L- lysine may also be used.

The present inventors also have shown that low doses of poly-lysine, either alone or in combination with monomeric lysine, produce a desirable reduction in kidney retention of radioactivity and cytotoxic agents. It also has been found that poly-lysine or poly-lysine/lysine combinations can be administered at lower dosages and less frequently while still retaining the desired reduction of kidney retention of radioactivity and cytotoxic agents. This leads to a lower overall dosage of amino acids to patients, reducing the risks of toxicity.

The present invention also is useful in therapeutic applications using protein conjugates such as antibody fragment conjugates, in which a cytotoxic agent is conjugated to the protein. Clearance of the protein conjugate through the kidney can lead to kidney damage caused by accumulation of the cytotoxic agent in the kidney. Administration of monomeric or polymeric lysine reduces the accumulation of the cytotoxic agent, and reduces kidney damage.

A. Selection and Labeling of Proteins and Antibodies

The protein conjugates of the present invention include, but are not limited to cytotoxic agent conjugates and radiolabeled conjugates of proteins of molecular weight greater than about 10 kD. The proteins include glycoprotein,

lipoprotein, or the like, e.g. hormones, lymphokines, growth factors, albumin, cytokines, enzymes, immune modulators and receptor proteins antibodies and antibody fragments. The protein conjugates of the present invention also includes peptides and polypeptides of molecular weight less than about 10 kD. Metabolic products of the above cytotoxic agent conjugates and the radiolabeled conjugates are also included within the scope of the present invention.

Cytotoxins conjugated to proteins in the methods of the present invention include, but are not limited to the group consisting of taxol, nitrogen mustards, ethylenimine derivatives, alkyl sulfonates, nitrosoureas, triazenes, folic acid analogs, pyrimidine analogs, purine analogs, vinca alkaloids, antibiotics, enzymes, platinum coordination complexes, substituted urea, methyl hydrazine derivatives, adrenocortical suppressants, hormones, antagonists, camptothecin, and endostatin, abrin, alpha toxin, diphtheria toxin, exotoxin, gelonin, pokeweed antiviral protein, ricin, saporin, and Onconase. The skilled artisan will be aware that additional cytotoxic agents can be used and are within the scope of the present invention.

The protein conjugates of the present invention may include a bifunctional linker which contains both a group capable of coupling to a cytotoxic agent or chelator and a group capable of coupling to the protein. The bifunctional linker can be coupled to the protein by means which are known in the art. For example, a linker containing an active ester moiety, such as an N-hydroxysuccinimide ester, can be used for coupling to lysine residues in the protein via an amide linkage. In another example, a linker containing a nucleophilic amine or hydrazine residue can be coupled to aldehyde groups produced by glycolytic oxidation of protein carbohydrate residues. In addition to these direct methods of coupling, the linker can be indirectly coupled to the protein by means of an intermediate carrier such as an aminodextran. In these embodiments the modified linkage is via either lysine, carbohydrate, or an intermediate carrier.

In a preferred embodiment, the linker is coupled site-selectively to free thiol residues in the protein. Moieties which are suitable for selective coupling to thiol groups on proteins are well known in the art. Examples include disulfide compounds, α -halocarbonyl and α -halocarboxyl compounds, and maleimides. When a nucleophilic amine function is present in the same molecule as an α -halo carbonyl or carboxyl group the potential exists for cyclization to occur via intramolecular alkylation of the amine. Methods to prevent this problem are well known to one of ordinary skill in the art, for example by preparation of molecules in which the amine and α -halo functions are separated by inflexible groups, such as aryl groups or trans-alkenes, that make the undesired cyclization stereochemically disfavoured.

It will also be appreciated that the chelators and cytotoxic agents of the present invention can be coupled directly to the particular targeting proteins.

The present invention can be used to reduce the kidney retention of radiolabeled protein conjugates. Radiolabeled conjugates are described in U.S. Patent 5,753,206 which is incorporated herein by reference in its entirety and includes antibody fragment conjugates that are radiolabeled by any means currently known or that may be known in the future. The term "antibody fragment" as used herein means a molecule which specifically binds to a complementary antigen and which is derived from a whole immunoglobulin by cleavage, by recombinant methods or by any other process that results in a functional equivalent of a conventional antibody fragment. Examples of suitable antibody fragments include divalent fragments, e.g., $F(ab)_2$, $F(ab')_2$, monovalent fragments, e.g., Fab, Fab', Fv, single chain recombinant forms of the foregoing, and the like. Antibody fragments may be glycosylated, for example containing carbohydrate moieties in the antibody variable regions.

The present invention can be used to reduce the kidney retention of antibody fragments that are radiolabeled with any radioisotope useful for RAID or RAIT. Examples of radioisotopes

that are useful for RAID applications are ^{99m}Tc and ^{111}In . Radioisotopes that are useful for RAIT include ^{32}P , ^{90}Y , ^{186}Re and ^{188}Re .

The antibody fragment may be radiolabeled by any method known to the art, or by any method that is discovered in the future. For a discussion of methods for radiolabeling antibody fragments see "Cancer Therapy with Monoclonal Antibodies", D.M. Goldenberg ed. (CRC Press, Boca Raton, 1995). The present invention is useful, therefore, for preventing kidney retention of antibody fragments that are labeled, *inter alia*, by conjugation of a radiometal-binding chelate, by direct radiometal labeling of hinge region sulfhydryl groups, or by radioiodination via the chloramine-T, iodogen, or Bolton-Hunter methods.

B. Administration of Lysine and Poly-lysine for Therapy and Diagnosis.

Generally, the overall dosage of lysine and/or poly-lysine administered to a patient will be that required to reduce the kidney retention of radiolabeled protein or antibody fragment to acceptable levels without causing lysine-induced toxicity. The dosage will vary depending upon such factors as the patient's weight, height, general medical condition, and previous medical history. Any physiologically acceptable salt of lysine or poly-lysine can be used, as can any physiologically acceptable carboxyl derivative of lysine or poly-lysine.

Administration of the monomeric or polymeric lysine or mixture thereof to a patient can be oral, intravenous, intraarterial, intraperitoneal, intramuscular, subcutaneous, or by perfusion through a regional catheter. Administration may be via single or multiple boluses, or by continuous or discontinuous infusion. Administration can also be by means of at least one injection of a bolus of a solution of said monomeric or polymeric lysine followed by oral administration in a physiologically acceptable carrier.

Equivalent effects on kidney uptake can be achieved with significantly lower doses of poly-lysine than is needed for monomeric lysine. Either D-, D/L- or L-poly-lysine, or any mixture thereof, may be used. The molecular weight range of the poly-lysine may be from 1-60 kDa, although higher molecular weight polymer could also be used. In a preferred embodiment the poly-lysine has a molecular weight range of approximately 1-4 kDa. In another preferred embodiment the poly-lysine has a molecular weight range of approximately 15-30 kDa.

In a preferred embodiment the dosage of poly-lysine administered to a patient in a single bolus is 1-50 mg. Repeated dosage or continuous infusion of poly-lysine is typically not required for diagnostic applications, but may be needed for prolonged therapeutic protocols. The concentration of poly-lysine in the solution is typically 1-50 g/l, though concentrations outside this range may also be used.

The present inventors have also found that use of mixtures of monomeric and polymeric lysine can reduce uptake of radioactivity and cytotoxic agents in the kidney to a similar or greater extent than monomeric lysine alone, but with less frequent administrations. Any combination of D- or L-lysine and D-, D/L, or L-poly-lysine may be used to achieve this effect. The molecular weight range of the poly-lysine used in the mixtures is preferably the same as when poly-lysine is used alone, as described *supra*. The dosage of monomeric lysine administered to a patient in the mixture is typically from 1-200 g, and that of polymeric lysine is typically from 1-50 mg. In both cases, however, a lower or higher dose may be used. In a preferred embodiment of the invention the dosage of monomeric lysine administered to the patient in a single bolus of the mixture is 2-35g and that of polymeric lysine is 5-25mg. The concentration of monomeric lysine in the solution is typically between 3-200 g/l, and that of polymeric lysine is 0.5-100 g/l. In a preferred embodiment the concentration of monomeric lysine

in the solution is between 2-35 g/l, and that of polymeric lysine is 10-25 g/l.

C. Determination of effects on kidney uptake of radioactivity and cytotoxic conjugates

Initial dosage ranges for achieving the desired reduction in kidney uptake of radioactivity and cytotoxic conjugates can be determined by using standard animal model systems. Animals are injected with protein conjugates and the level of kidney uptake is compared in animals that are treated with different amounts of lysine or lysine-containing compounds. The results obtained are compared to the results obtained in control animals which receive only the protein conjugates. Observations concerning the effect on the reduction of kidney uptake of radiolabeled conjugates and conjugates of cytotoxic agents can generally be made by routine histological examinations of the kidney using methods that are well-known in the art. Another method of assaying kidney toxicity is to measure the reduction or elimination of blood in the urine.

A preferred model system is the mouse. For determining the reduction of kidney uptake in non-tumor-bearing subjects the preferred mouse strain is BALB/c mice (Charles River Laboratories, Boston, MA). The preferred strain for determining reductions in tumor-bearing subjects is athymic mice (Harlan, Madison, WI), carrying an engrafted human tumor, such as that produced by subcutaneous injection of the human colon carcinoma cell line GW39 (Primus et al., *Cancer Res.* 33:2977 (1973)). Mice are injected intravenously into the tail vein with approximately 5-10 μ g of radiolabeled antibody fragment protein. Typical doses of radioactivity are 25-40 μ Ci of ^{99m}Tc , ^{188}Re , or ^{111}In , 4 μ Ci of ^{88}Y , and 10 μ Ci of ^{125}I . For example, mice are sacrificed at predetermined time periods after injection of the labeled antibody fragments, for example at 4 h and 24 h for ^{99m}Tc , and at 4, 24, 72, 96 and 168 h for ^{125}I , ^{111}In , ^{88}Y and ^{188}Re -labeled antibodies. The mice are sacrificed and dissected and the

amount of activity in the kidney is determined by standard methods. An injection standard is used to correct for radioactive decay.

Comparison of the results in the control mice with those in the mice that receive the lysine and/or poly-lysine allows determination of the optimal dosage required to achieve the desired effect on kidney uptake of radioactivity. Determination of the dosage of lysine and/or poly-lysine that can be tolerated before toxic effects are observed can be achieved by physiological examination and histological observation of organs by methods. The optimal dosages of lysine and/or poly-lysine that are determined using the mouse model is used to guide the determination of appropriate dosage levels in human subjects using a standard dosage escalation regimen. Methods for determining effective dosages of lysine and/or poly-lysine for reducing kidney uptake are further illustrated in the Examples, below.

Kits containing RAID and RAIT reagents can advantageously include monomeric and/or polymeric lysine.

The present invention, thus generally described, will be understood more readily by reference to the following examples, which are provided by way of illustration and are not intended to be limiting of the present invention.

Examples

General Methods

Preparation of Radiolabeled Antibody Fragments

The murine monoclonal anti-carcinoembryonic antigen (anti-CEA) antibody NP-4 and the second generation antibody anti-CEA MN-14 have been described previously (Sharkey *et al.*, *Cancer* 71:2082 (1993). The anti-colon-specific antigen-p (CSAp) antibody Mu-9 and the anti-B-cell lymphoma (anti-CD22) antibody LL2 (EPB2) have also been reported. See Blumenthal *et al.*, *Cancer Immunol. Immunother.* 32:303 (1991) and co-pending U.S. Application Serial No. 08/289576. Antibodies were purified from mouse ascites fluid by standard methods using protein A and ion-exchange chromatography on S- and Q-Sepharose (Pharmacia, Piscataway, NJ) at 4°C. Purity was ascertained by immunoelectrophoresis, sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS-PAGE) and isoelectric focusing.

All antibody fragments were prepared by standard methods. F(ab')₂ fragments of the NP-4, Mu-9 and LL2 antibodies were prepared by pepsin digestion, and F(ab)₂ fragments of MN-14 were generated by papain digestion. The fragments were purified by protein A chromatography on Protein A-Sepharose (Pharmacia), using the conditions recommended by the manufacturer. Fab' fragments of NP-4, LL2 and Mu-9 were generated from the corresponding F(ab')₂ fragments, and the Fab fragment of MN-14 was generated from the corresponding F(ab)₂ fragments by reduction with cysteine using standard methods.

Fab' fragments NP-4 and LL2 were prepared for ^{99m}Tc-labeling by following the method described in U.S. Patent 5,061,641. For ¹⁸⁸Re-labeling, Fab' fragments of Mu-9 and LL2 containing free sulfhydryl groups were prepared by reduction with 2-20 μM 2-mercaptoethanol for 10 min at 4°C. The reaction products were purified by gel filtration chromatography, mixed with stannous tartrate in 50 mM PBS, pH 5.3, lyophilized and stored under an inert atmosphere.

For ^{111}In and ^{88}Y -labeling, isothiocyanate benzyl-DTPA (SCN-Bz-DTPA) conjugates of the F(ab)_2 and Fab fragments of MN-14, and the F(ab')_2 fragment of LL2 were prepared by adding SCN-Bz-DTPA to the antibody (5.0 mg/ml), previously dialyzed against 100 mM Hepes buffer, pH 8.6, containing 150 mM NaCl, at a 8:1 molar excess of DTPA to MAb. After overnight incubation at room temperature, the antibody conjugates were purified from unreacted SCN-Bz-DTPA by gel filtration chromatography on a 1 x 50 cm column of Sephadex G-50 (Pharmacia).

Technetium-99m was obtained from a $^{99}\text{Mo}/^{99\text{m}}\text{Tc}$ generator system (Syncor, Fairfield, NJ) as a solution of sodium pertechnetate in 0.9% sodium chloride. Rhenium-188 was obtained from a tungsten-188 ($^{188}\text{W}/^{188}\text{Re}$) generator system (Oak Ridge National Laboratory, Oak Ridge, TN) as a solution in 0.9% sodium chloride. Indium-111 was purchased as $^{111}\text{InCl}_3$ in 0.1M HCl from NEN-DuPont (Boston, MA). Yttrium-88 was obtained as $^{88}\text{YCl}_3$ in 6 M HCl from NEN-DuPont. Iodine-125 was obtained from NEN-DuPont.

Radioiodination was carried out with ^{125}I using the iodogen method as previously described. See Fraker et al., *Biochem. Biophys. Res. Comm.* 80:849 (1978). For $^{99\text{m}}\text{Tc}$ - and ^{188}Re -labeling, the lyophilized antibody fragments prepared as described above were reconstituted with sodium pertechnetate or perrhenate in 0.9 % saline. Labeling with ^{111}In and ^{88}Y was carried out by standard methods. See, for example, "Advanced Methods for Radiolabeling Monoclonal Antibodies with Therapeutic Radionuclides" in Goldenberg, *supra*.

Determination of Reduction of kidney uptake of radioactivity

Female BALB/c mice, 19 to 22 g of weight, 4 to 5 weeks of age (Charles River Laboratories, Boston, MA) were used as non-tumor-bearing animals. For a human tumor xenograft model, the human colon carcinoma cell line GW39 (Primus et al. *supra*) was grown subcutaneously in 5- to 6 week-old female athymic mice (Harlan, Madison, WI). Animals were injected intravenously into the tail vein with approximately 5-10 μ g of antibody fragment protein (i.e., 25-40 μ Ci of ^{99m}Tc , ^{188}Re , and ^{111}In , resp.; 4 μ Ci of ^{88}Y , and 10 μ Ci of ^{125}I , respectively).

The mice were sacrificed after 4 h and 24 h for ^{99m}Tc -labeled antibodies, and after 4, 24, 72, 96 and 168 h for ^{125}I , ^{111}In , ^{88}Y and ^{188}Re -labeled antibodies. Mice were sacrificed by anesthesia with sodium pentobarbital and bled by cardiac puncture. After cervical dislocation, the animals were dissected. Radioactivity in the tumors and tissues (liver, spleen, kidney, lung, intestine, blood and bone) was determined by gamma scintillation counting using an injection standard to correct for physical decay.

For determination of whole-body clearance and organ uptake kinetics in humans, the ROI (region of interest) technique was applied onto whole-body scans obtained at 10 min, 1 h, and 24 h after antibody administration (Siemens BODYSCAN, Siemens Gammasonics, Erlangen, FRG). Geometric means were calculated, and after correction for radioactive decay, all values were expressed in percent of injected dose by referring to the whole-body counts at 10 min p.i. without previous voiding of the urinary bladder (by definition 100% of the injected dose).

Preparation of amino acid solutions

L-lysine, D-lysine and L-arginine monohydrochloride salts (Sigma, St. Louis, MO) were dissolved in phosphate-buffered saline (PBS) at a concentration of 160 mg/ml. L-lysine ethyl

ester dihydrochloride was dissolved in PBS to yield a concentration of 80 mg/ml. Poly-L-lysine (1-4 kDa and 15-30 kDa) were used as hydrochloride salts at concentration of 25 mg/ml and 10 mg/ml, respectively.

A commercially available amino acid solution (Periamin X[®], Pfrimmer -Kabi-Pharmacia, Erlangen, FRG, containing 8.2 g L-lysine/l and 12 g L-arginine/l.), intended for parenteral nutrition of patients was used as an amino acid source in human patients.

Example 1. Reduction in kidney uptake by lysine and/or poly-lysine

Administration of lysine and poly-lysine-containing solutions

Animals were injected with the lysine and/or poly-lysine solutions intraperitoneally in amounts and at frequencies as shown below. Human patients were infused intravenously over a three-hour period with 1.5 liters of a commercially available amino acid solution starting 15 min before the antibody injection. The data of a total of 77 patients examined with ^{99m}Tc-labeled anti-CEA Fab' (NP-4: n=19; F023C5: n=58) were taken as controls (infused with the same volume of 0.9 % saline instead of the amino acid solution).

Example 2. Effect of L-lysine on Kidney Uptake

A. In mice

Typical kidney uptake and organ kinetics of radiolabeled Fab' and F(ab')₂ fragments are shown in Figure 1. ^{99m}Tc-labeled Fab' fragments showed a rapid renal uptake, reaching a maximum at 2 h after intravenous antibody injection. At this time-point a plateau was reached, lasting approximately until 4-6h p.i., after which excretion of originally retained activity predominated. Peak values of kidney uptake ranged from 65.0 ± 10.9 %ID/g to 120 ± 15% %ID/g, accounting for an absolute

uptake in both kidneys of in between 18 and 35 percent of the originally injected activity.

F(ab')₂ - kinetics With Y and In labeled F(ab)₂ there is a continuous uptake up to 24 h p.i., reaching its highest value at about 55% of the injected dose /g for In, and about 43% of the injected dose /g for the Y label, with subsequent wash-out.

Figure 2 shows the dose-effect relationship between L-lysine, administered i.p., and the kidney uptake of ^{99m}Tc-labeled Fab' fragments of the monoclonal anti-CEA antibody NP-4 in BALB/c mice. L-lysine was injected i.p. in hourly intervals from 30 min before until 3 h after intravenous antibody injections (-30', 1 h, 2 h, 3 h p.i.). Doses below four times 100 µg/g body weight (given in hourly intervals), produced no significant effect could be observed. At doses above this threshold, a strong dose-effect relationship was found. At 4 x 2000 µg/g, kidney uptake was reduced to 19 ± 1% of the untreated control group at 4 h p.i. The maximum tolerated dose (MTD) of L-lysine in mice was reached at 4 x 2500 µg/g (injected i.p. in hourly intervals). This dose was tolerated without any obvious short or long term toxicity (monitored for three months). At higher doses, the mice began to develop fluid in the body cavities (e.g. pleural effusions). No additional effect on kidney uptake was observed in lysine doses above this MTD (see Figure 2).

The effect of uptake reduction of ^{99m}Tc-Fab' persisted for 24 hours. This effect was obscured somewhat because in the control group 50-70% of the originally retained technetium had been excreted at this time-point, and therefore lysine-induced reduction appeared less profound than at earlier time-points.

In nude mice bearing human colonic xenografts, no significant influence on the tumor uptake or uptake in any other organ than the kidneys was seen by the lysine treatment (see Table 1).

Less frequent L-lysine administration (once 30 min before antibody administration or twice 30 min before and 1 h after

antibody injection) was significantly less effective than the four injections in hourly intervals. For example, a single injection at 30 min. before antibody administration reduced kidney uptake of Tc-Fab' only to 56.4 % of the control amount, and two injections (at 30 min. before and 1 h after antibody) reduced kidney uptake to 44.1 % of the control amount. In comparison, four injections reduced kidney uptake to 15.5 % of control. See Table 2.

L-lysine had comparable effects on ^{188}Re -labeled Fab' fragments of the anti-CSAp antibody Mu-9. See Figure 3. Lysine treatment reduced kidney uptake to 29 % of the untreated control after 4 h p.i. No effect on the uptake in the tumor or other organs was observed. Table 3 summarizes the dosimetry of ^{188}Re -Mu-9 Fab' under control and lysine treatment conditions. The kidney dose under lysine-treatment is approximately 30 % of the renal dose in the control group, whereas no effect on the tumor dose could be observed.

To test the effectiveness of L-lysine in reduction of kidney uptake of indium and yttrium-labeled Fab' fragments, a mixture of ^{111}In and ^{88}Y -labeled Fab' fragments of the anti-CEA antibody MN-14 were injected into GW39 colon carcinoma bearing nude mice (injected with four times 2000 $\mu\text{g/g}$ L-lysine; the control group were untreated). Figure 4 and Table 4 summarize the results.

Lysine also reduced the kidney uptake of Fab' fragments labeled with isotopes that are not retained significantly by cells (e.g. iodine). Table 5 summarizes the organ kinetics of an ^{125}I -labeled Fab' fragment of NP-4. Although the kidney uptake of iodine-labeled Fab' was found much lower than with all radiometals, it was still reduced by L-lysine to 38.7 % of the nontreated control at 4 h p.i., which results in a 50 % reduction of the absorbed dose to the kidney, calculated for a ^{131}I -labeled Fab' fragment.

Figure 6 and Table 6 summarize the reduction of kidney uptake observed with F(ab')_2 fragments. Uptake with ^{111}In -and ^{88}Y -

$F(ab')_2$ was reduced to approximately 30 % of control by lysine treatment. As was observed with Fab' fragments, reduction of uptake persisted for several days. See Figure 6.

Table 1

Dose-effect relationship of treatment with L-lysine and poly-L-lysine (1 - 4 kD) in GW39 colonic cancer xenograft bearing nude mice with anti-CEA ^{99m}Tc-Fab' NP-4

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control	L-lysine	poly-L-lys
0.5mg/g lys	2mg/g lys	0.15mg/g poly
injection i.p. at	injection i.p. at	i.v. at
-30', 1, 2, 3 h	-30', 1, 2, 3 h	-30',

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4 h p.i.

GW39	3.5±0.6	3.2±0.5	5.0±1.9	3.0±1.0
liver	2.0±0.3	1.6±0.2	1.8±1.1	1.5±0.2
spleen	1.3±0.1	1.0±0.1	1.3±0.1	1.0±0.1
kidney	58.9±9.6	37.1±3.7	17.5±4.3	30.8±6.0
% of contr.	100.0	63.0	29.7	52.2
lung	3.1±0.2	5.4±1.3	4.5±1.4	3.1±1.2
blood	1.7±0.3	1.6±0.2	1.9±0.2	1.7±0.2

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24 h p.i.

GW39	1.7±0.2	1.9±0.5	2.2±0.6	2.2±0.3
liver	1.0±0.1	0.9±0.2	1.4±0.3	0.9±0.1
spleen	0.6±0.1	0.5±0.3	0.7±0.3	0.7±0.1
kidney	26.7±3.6	18.7±3.9	8.0±3.2	25.3±5.7
% of contr.	100.0	70.0	30.0	94.7
lung	0.3±0.1	0.4±0.2	0.9±0.6	0.8±0.6
blood	0.3±0.0	0.5±0.3	0.3±0.1	0.3±0.0

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Table 2

Effect of less frequent administration of L-lysine on the biodistribution of ^{99m}Tc -Fab' NP-4
 5 in BALB/c mice (uptake values in % ID/g at 4 h p.i.)

organ	control -30'	L-lysine (2000 $\mu\text{g/g}$)			
		injection once at -30' and 1h	injection twice at -30', 1, 2, 3h		
15	liver	2.5 \pm 0.3	4.9 \pm 2.4	2.7 \pm 0.5	2.2 \pm 0.5
	spleen	2.3 \pm 0.2	3.2 \pm 1.2	2.0 \pm 0.2	1.4 \pm 0.4
	kidney	122.6 \pm 20.8	69.1 \pm 9.8	54.1 \pm 6.0	19.0 \pm 5.7
	% of contr.	100.0	56.4	44.1	15.5
	lung	3.4 \pm 0.2	6.2 \pm 3.8	4.6 \pm 1.7	2.1 \pm 0.9
20	blood	3.3 \pm 0.4	4.3 \pm 1.8	3.0 \pm 0.6	2.4 \pm 0.4

Table 3

Dosimetry of $^{188}\text{Re-Fab'}$ Mu-9

5

	control		L-lysine		
	cGy/mCi	T/NT	cGy/mCi	T/NT	lys/contr.
10					
GW39	620.3		763.4		1.23
liver	611.3	1.01	587.6	1.30	0.96
15					
spleen	306.0	2.03	289.0	2.64	0.94
kidney	7818.8	0.08	3085.2	0.25	0.39
20					
lung	537.7	1.73	410.4	1.86	0.76
blood	256.4	2.42	203.1	3.76	0.79
bone	120.9	5.13	107.3	7.11	0.89

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Table 4

Effect of treatment with L-lysine (four times 2000 µg/g i.p.) on the dosimetry of anti-CEA Fab' MN-14 in GW39 colonic cancer xenograft bearing nude mice. A mixture of ¹¹¹In and ⁸⁸Y-labeled Bz-DTPA-Fab' was injected. The ⁹⁰Y-dosimetry was calculated based on the ⁸⁸Y and ¹¹¹In biodistribution separately.

		control cGy/mCi based on ¹¹¹ In	T/NT yttrium	L-lysine cGy/mCi based on ¹¹¹ In	T/NT yttrium
10	GW39	5668.8	4953.8	6478.1	5751.1
	liver	1736.1	1339.9	2096.8	1651.9
	spleen	607.6	626.9	564.5	499.2
20	kidney	34245.3 100%	33057.2 100%	7849.0 22.9%	6231.0 18.8%
	lung	501.0	385.6	662.8	431.1
	blood	751.2	493.2	508.7	588.4
	intestine	737.1	519.9	768.1	560.2
30	bone	391.2	493.2	508.7	588.4
	washed bone	187.0	396.7	277.6	486.8

Table 5

Effect of L-lysine (4x2mg/g) on the kidney uptake of ^{125}I -labeled Fab' NP-4 and the dosimetry for a ^{131}I -labeled Fab'

	organ	^{131}I -Fab' NP-4		
		control cGy/mCi	L-lysine cGy/mCi	lys/control
10				
	liver	81.9	77.2	0.94
15	spleen	51.8	42.1	0.81
	kidney	229.5	116.6	0.51
	lung	126.4	120.2	0.95
20	blood	117.7	116.6	0.99

Table 6

Effectiveness of L-lysine on the kidney uptake and the dosimetry of yttrium-labeled F(ab')₂ fragments of the anti-CD22 antibody LL2

5

organ	%ID/g 4 h p.i.		%ID/g 48 h p.i. dose		(cGy/mCi)	
	contr.	lys	contr.	lys	contr.	lys
=====						
liver	5.9±0.5	8.8±1.6	4.0±0.2	4.5±0.5	4509.8	5375.0
spleen	3.1±0.3	3.5±0.7	3.9±0.5	3.8±0.6	2989.2	3031.9
kidney	22.9±2.8	13.4±0.8	26.2±4.3	13.0±1.6	21230.7	10813.9
% control	100.0	58.5	100.0	49.6	100.0	50.9
lungs	5.2±0.9	6.9±2.4	1.9±0.3	1.9±0.2	1777.8	2192.1
blood	13.4±2.4	12.8±4.7	1.5±0.2	1.4±0.2	4146.4	3651.3

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B. In humans

Five patients undergoing radioimmunodetection studies with ^{99m}Tc -labeled Fab' fragments of two anti-CEA antibodies were infused over a three-hour period with a commercially available amino acid solution, containing a total of 1.75 g of L-lysine and 3.75 g of L-arginine as basic amino acids. Quantification of organ uptake kinetics showed a decrease of kidney uptake. See Figure 7. This effect was more pronounced at 24 than at 4 h p.i. Whole-body retention showed slightly lower values in the treated patients.

Example 3. Effect of D-lysine on Kidney Uptake

D-lysine was tested under the same conditions as L-lysine (four hourly injections i.p.). D-lysine was found to be equally as effective as the L-isomer in lowering kidney uptake.

Example 4. Effect of polymeric lysine compounds

To attempt to reduce the necessity of frequent injections of the kidney uptake reducing agent, polymeric substances of higher molecular weight were investigated. The results obtained are summarized in Table 7. Poly-L-lysine with a molecular weight range of 1-4 kDa reduced kidney uptake with a single i.p. injection at significantly lower doses than the monomer. However, the MTD was reached at 300 $\mu\text{g/g}$, so that the possible maximal absolute uptake reduction was less than with the monomer. The potency of poly-L-lysine increased with increasing molecular weight. L-lysine of molecular weight 15-30 kDa showed 50 % uptake reduction at 1 x 20 $\mu\text{g/g}$ (MTD 20 $\mu\text{g/g}$).

Table 7

Effect of poly-L-lysine of different molecular weight on the biodistribution of ^{99m}Tc -Fab' NP-4 at 4 h p.i.

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organ	control	poly-L-lysine administered once 30' before antibody			
		100 $\mu\text{g/g}$	200 $\mu\text{g/g}$	300 $\mu\text{g/g}$	10 $\mu\text{g/g}$ 15-30kD
liver	3.4 \pm 0.4	3.0 \pm 0.2	3.0 \pm 0.4	3.5 \pm 0.8	7.3 \pm 3.7
spleen	2.1 \pm 1.1	3.0 \pm 0.4	2.7 \pm 0.4	3.5 \pm 0.4	3.7 \pm 0.4
kidney	127.7 \pm 15.7	95.5 \pm 4.3	80.5 \pm 6.9	67.9 \pm 7.5	73.9 \pm 7.2
% control	100.0	74.8	63.0	53.2	57.8
					43.6
lungs	7.7 \pm 1.3	5.9 \pm 3.1	3.9 \pm 2.6	5.6 \pm 1.0	6.8 \pm 4.3
blood	5.1 \pm 0.4	3.8 \pm 0.1	4.4 \pm 0.5	5.3 \pm 1.4	4.7 \pm 0.3

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4.6 \pm 0.7

15

3.8 \pm 0.3

20

4.7 \pm 0.2

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poly-L-lysine administered twice (30' before and 1h after antibody)

kidney	73.0 \pm 6.0	43.7 \pm 10.0	41.6 \pm 8.8
% control	100.0	59.9	56.9

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Example 5. Effect of mixtures of monomeric and polymeric lysine compounds

To reach maximal effectiveness with as few injections as possible, combinations of lysine and poly-lysine were tested. The results obtained are summarized in Figure 5. Optimal results were found with two injections of a mixture of 2 mg/g lysine and 20 µg/g poly-lysine (15-30 kDa) at 30 min. before and 1 h after antibody administration. Despite the fact that both compounds were given close to their individual maximum tolerated dose, no acute or long-term toxicities could be noticed during monitoring for three months after treatment.

Example 6. Diagnostic Imaging of Lymphoma

A posterior abdomen scan of a patient with a gastric carcinoma in the antral region and liver metastasis in the left hepatic lobe who underwent RAID with ^{99m}Tc Fab' fragment of FO23C5 (Sorin Biomedical, Saluggia, Italy) and was infused with Periamin (see above). The kidney image obtained at 4h and 24h p.i. was much less intense than usually observed with fragments and therefore the primary tumor, which was adjacent to the left kidney, was readily visualized.

The invention has been disclosed broadly and illustrated in reference to representative embodiments described above. Those skilled in the art will recognize that various modifications can be made to the present invention without departing from the spirit and scope thereof.

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What is claimed is:

1. A method of reducing kidney retention of a protein conjugate in a patient, comprising administering to said patient one or more compounds selected from the group consisting of D-lysine, poly-D-lysine having a molecular weight in the range 1-60 kD, poly-L-lysine having a molecular weight in the range 1-60 kD, pharmaceutically acceptable salts thereof and carboxyl derivatives thereof,

wherein the pharmaceutically acceptable salts and carboxyl derivatives of poly-D-lysine or poly-L-lysine have a molecular weight in the range 1-60 kD,

whereby said compound or compounds reduce kidney retention of said conjugates.

2. A method according to claim 1, wherein said protein conjugate is selected from the group consisting of protein conjugates, peptide conjugates, polypeptide conjugates, glycoprotein conjugates, lipoprotein conjugates, antibody conjugates, antibody fragment conjugates and the metabolic products thereof.

3. A method according to claim 1, wherein said protein conjugate is a radiolabeled conjugate.

4. A method according to claim 3, wherein the radiolabel in said radiolabeled conjugate is an imaging isotope.

13. The method according to claim 1, wherein said compound is parenterally administered to said patient in a physiologically acceptable aqueous solution.

14. The method according to claim 13, wherein said physiologically acceptable aqueous solution is administered to said patient by continuous infusion.

15. The method according to claim 13, wherein said physiologically acceptable aqueous solution is administered to said patient by means of at least one injection of a bolus of said solution.

16. The method according to claim 15, wherein said physiologically acceptable aqueous solution is administered to said patient by means of at least one injection of a bolus of said solution followed by oral administration in a physiologically acceptable carrier

17. The method according to claim 1, wherein said compound is orally administered to said patient in a physiologically acceptable carrier.

18. A method of reducing kidney retention of a protein conjugate in a patient undergoing treatment with a targeting protein conjugate comprising administering to said patient, one or more compounds selected from the group

consisting of D-lysine, poly-D-lysine having a molecular weight in the range 1-60 kD, poly-L-lysine having a molecular weight in the range 1-60 kD, pharmaceutically acceptable salts thereof and carboxyl derivatives thereof,

wherein the pharmaceutically acceptable salts and carboxyl derivatives of poly-D-lysine or poly-L-lysine have a molecular weight in the range 1-60 kD,

whereby said compound or compounds reduce kidney retention of said conjugates.

19. A method according to claim 18, wherein said protein conjugate is selected from the group consisting of protein conjugates, peptide conjugates, polypeptide conjugates, glycoprotein conjugates, lipoprotein conjugates, antibody conjugates, antibody fragment conjugates and the metabolic products thereof.

20. A method according to claim 18, wherein said targeting protein conjugate comprises a ribonucleic acid binding protein.

21. A method according to claim 20, wherein said ribonucleic acid binding protein is a ribonuclease.

22. A method according to claim 21, wherein said ribonuclease is an onconase or recombinant form thereof.

23. A method according to claim 18, wherein said protein conjugate is a radiolabeled conjugate.

24. A method according to claim 22, wherein the radiolabel in said radiolabeled conjugates is an imaging isotope.

25. A method according to claim 22, wherein the radiolabel in said radiolabeled conjugates is a therapeutic isotope.

26. A method according to claim 18, wherein said protein conjugate is selected from the group consisting of radiolabeled hapten conjugates and haptens conjugated to a cytotoxic agent.

27. A method according to claim 18, wherein said protein conjugate comprises a cytotoxic agent.

28. The method according to claim 18, wherein D-lysine is administered to said patient.

29. The method according to claim 18, wherein poly-D-lysine is administered to said patient.

30. The method according to claim 18, wherein poly-L-lysine is administered to said patient.

31. The method according to claim 18, wherein a mixture of at least two of said compounds is administered to said patient.

32. The method according to claim 18, wherein said poly-D-lysine and said poly-L-lysine each have a molecular weight of 15-30 kD.

33. The method according to claim 18, wherein said compound is parenterally administered to said patient in a physiologically acceptable aqueous solution.

34. The method according to claim 33, wherein said physiologically acceptable aqueous solution is administered to said patient by continuous infusion.

35. The method according to claim 34, wherein said physiologically acceptable aqueous solution is administered to said patient by means of at least one injection of a bolus of said solution.

36. The method according to claim 35, wherein said physiologically acceptable aqueous solution is administered to said patient by means of at least one injection of a bolus of said solution followed by oral administration in a physiologically acceptable carrier .

37. The method according to claim 18, wherein said compound is orally administered to said patient in a physiologically acceptable carrier.

Abstract

Kidney uptake of antibody fragment conjugates and protein conjugates in patients is reduced by administration to the patient of one or more compounds selected from the group consisting of D-lysine, poly-D-lysine, or poly-L-lysine, or pharmaceutically acceptable salts or carboxyl derivatives thereof.

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Organ and kidney uptake kinetics

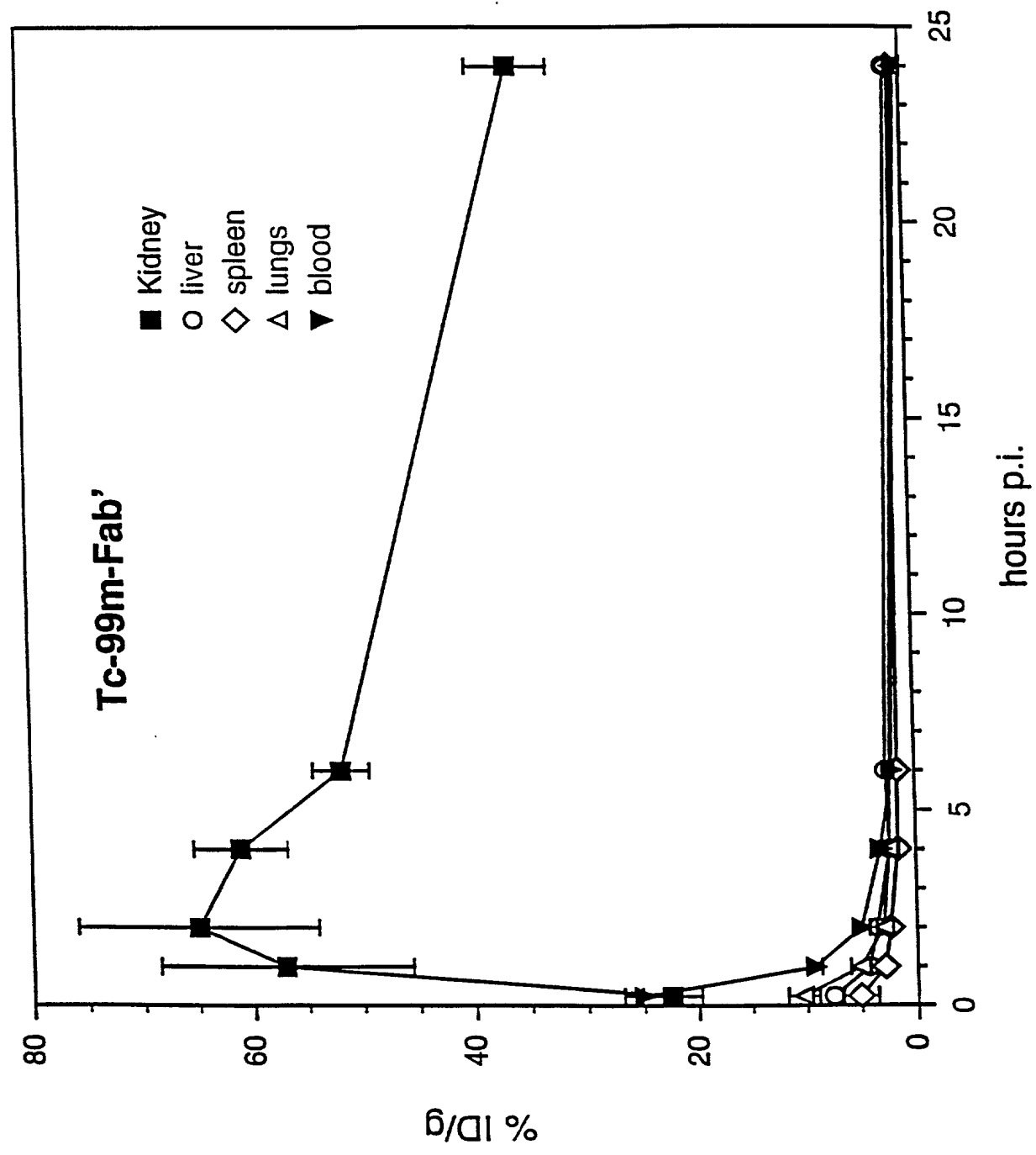


FIGURE 1

Kidney uptake reduction through L-lysine

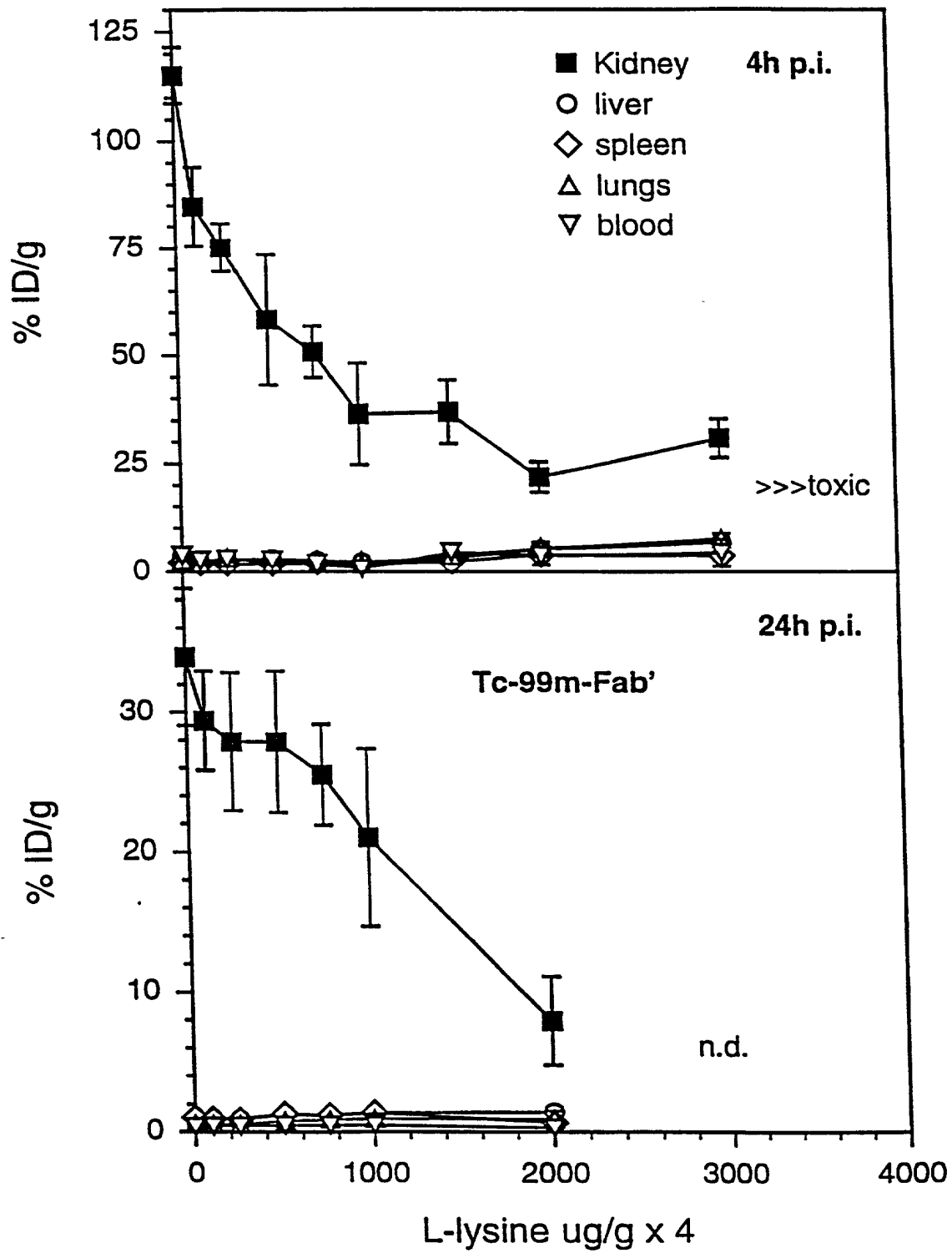


FIGURE 2

Re-188-Fab' Mu-9 in GW39 nude mice

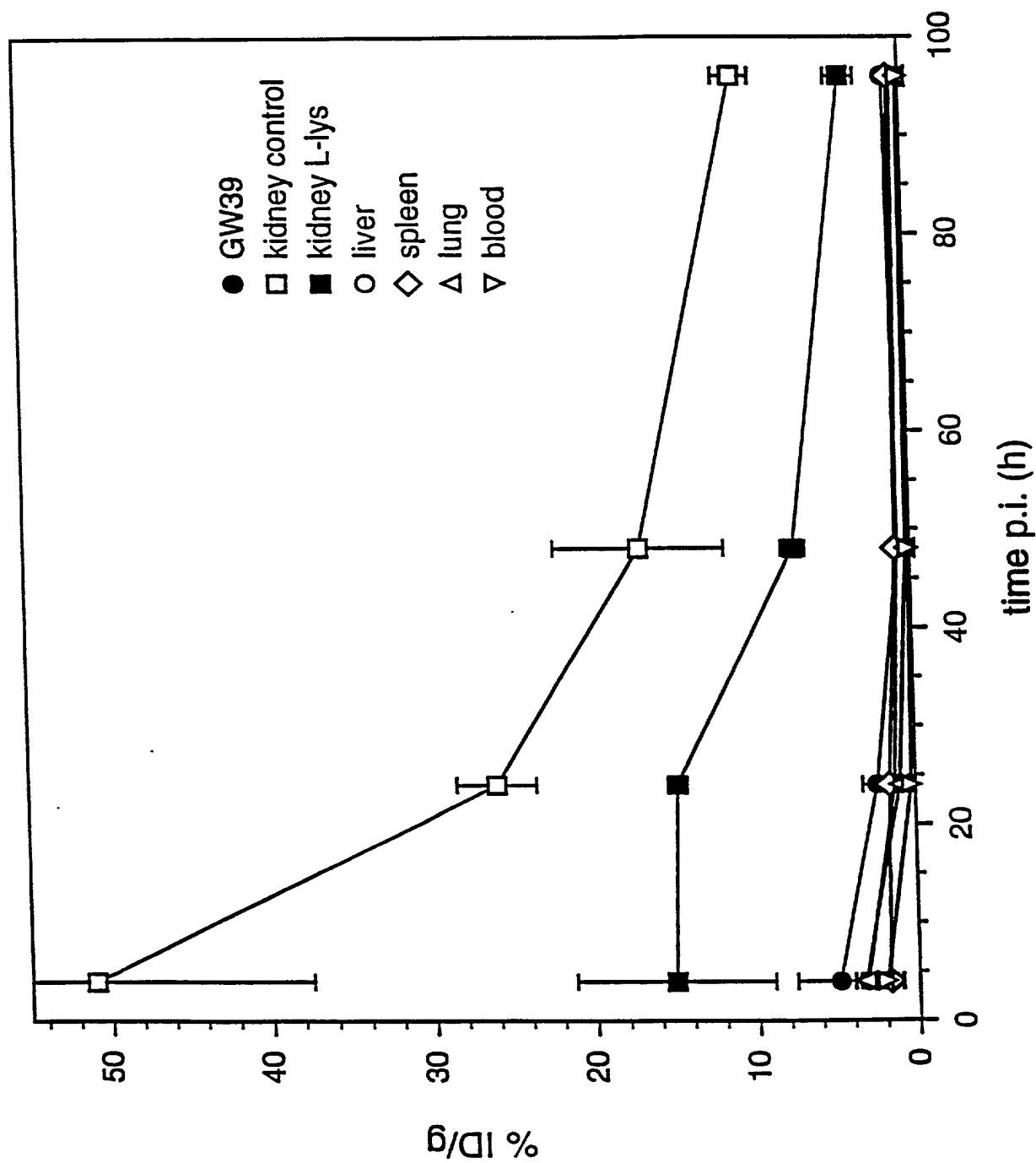


FIGURE 3

Indium and yttrium-BzDTPA-Fab' MN-14

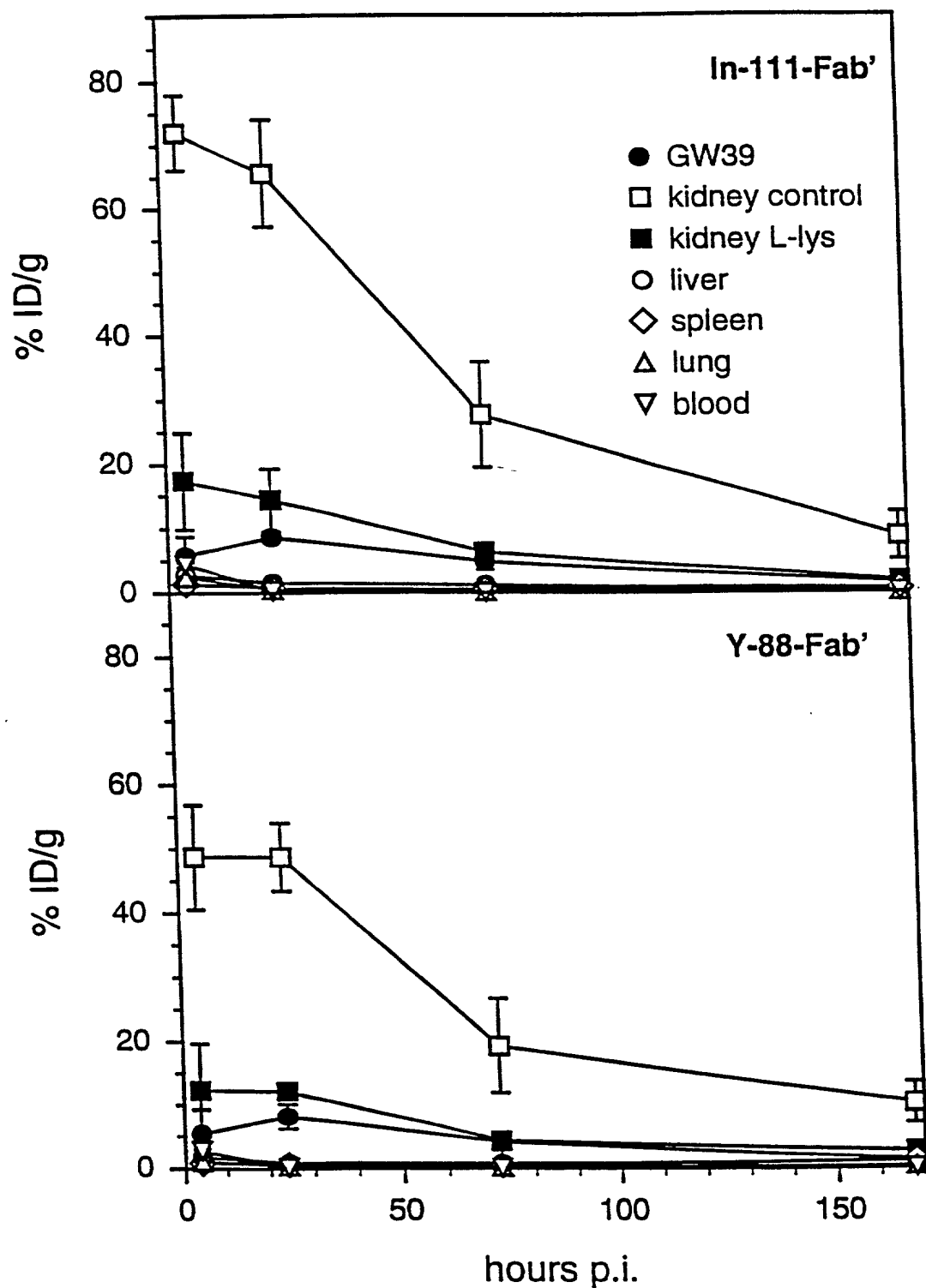


FIGURE 4

Effect of frequency of administration

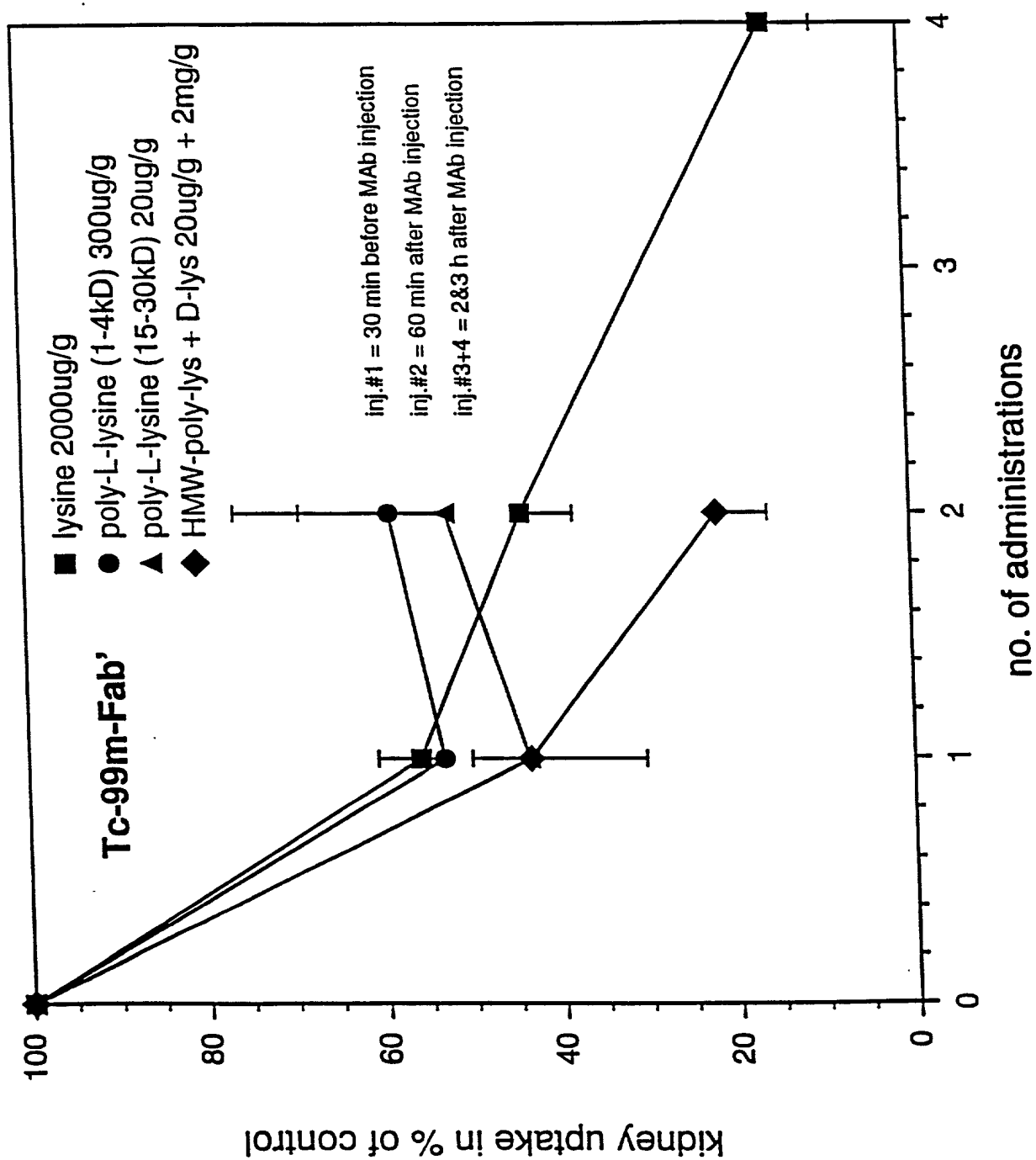


FIGURE 5

Indium and yttrium-BzDTPA-F(ab)2 MN-14

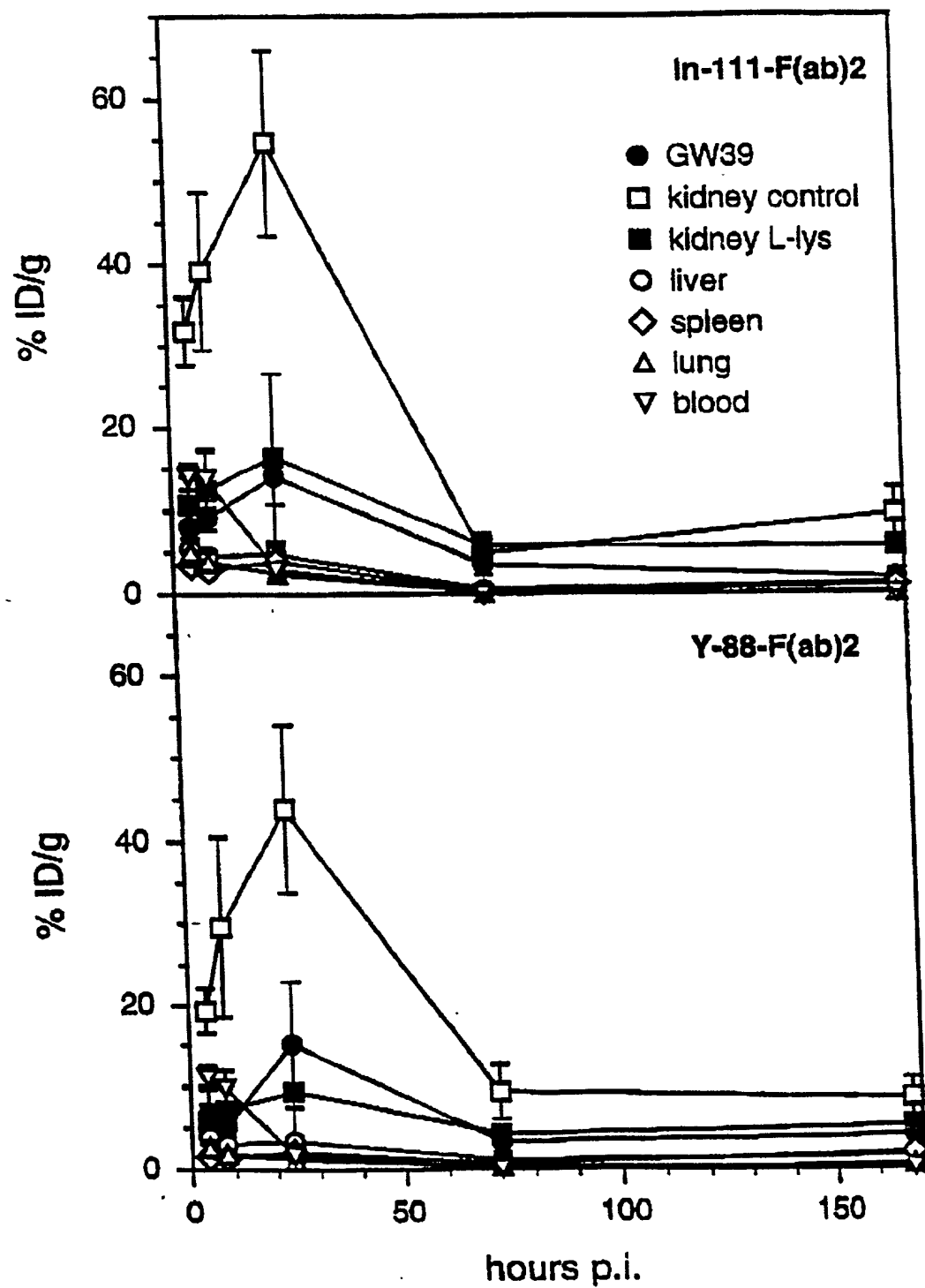


FIGURE 6

Whole-body and organ kinetics with Tc-99m-Fab'

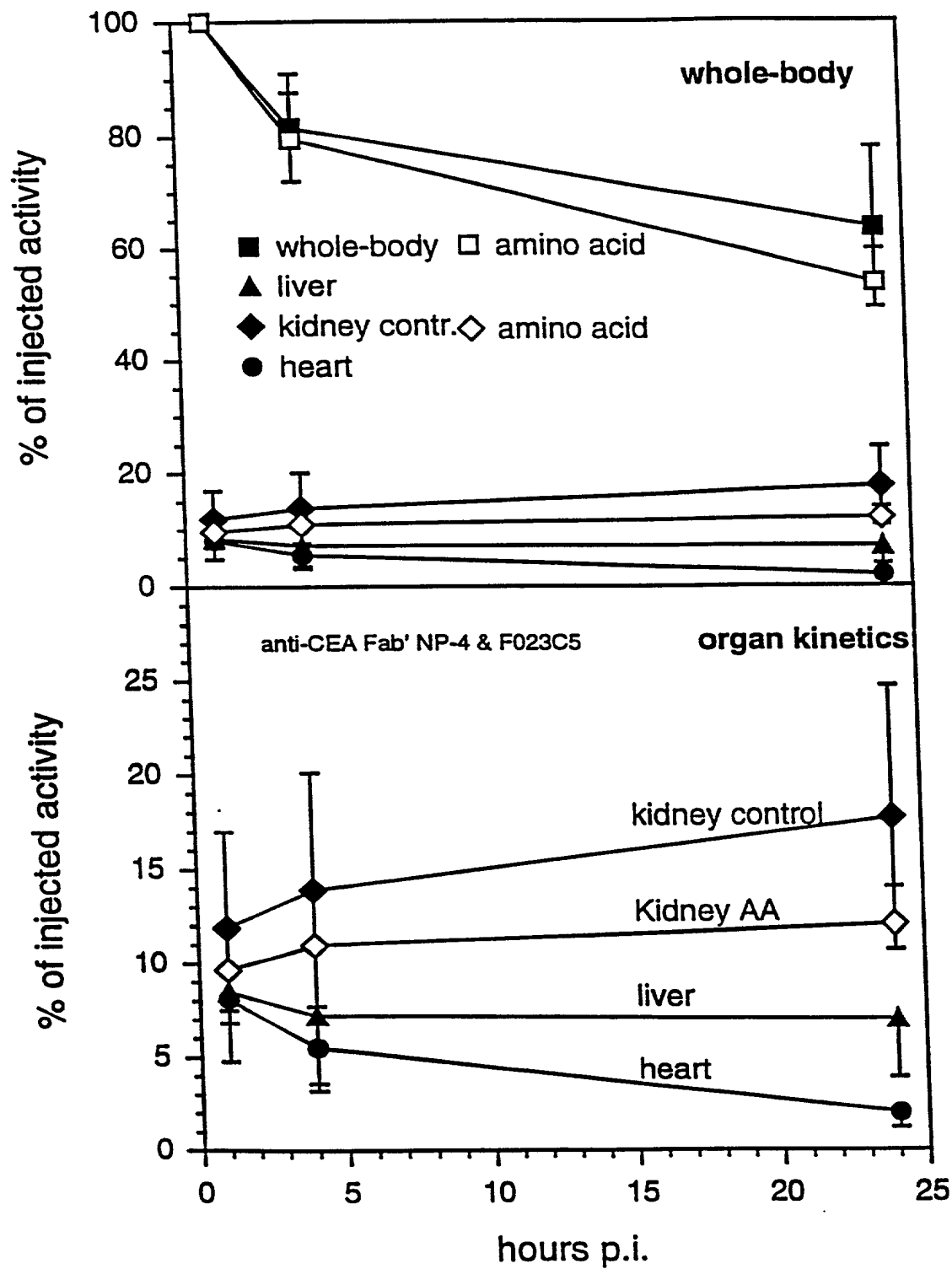


FIGURE 7